

T.C.  
ERCIYES ÜNİVERSİTESİ  
BİLİMSEL ARAŞTIRMA PROJELERİ  
KOORDİNASYON BİRİMİ

**PROJE BAŞLIĞI**

**Proje No:TA-05-19**

Bilimsel Araştırma Projesi

**SONUÇ RAPORU**

**Proje Yürütücüsü:**

Yard.Doç.Dr. Çetin Saatci  
Tıp Fakültesi\ Tıbbi Genetik AD

Prof.Dr.Yusuf Özkul  
Tıp Fakültesi\ Tıbbi Genetik AD



## İÇİNDEKİLER

	Sayfa No
ÖZET	4
ABSTRACT	5
1. GİRİŞ	6
2.GENEL BİLGİLER	7
3. GEREÇ VE YÖNTEM	9
4. BULGULAR	10
5.TARTIŞMA VE SONUÇ	11
6.KAYNAKLAR	12
7.EKLER	15

## ÖZET

Multiple myeloma hastalarının tedavi öncesi ve sonrası rezidual myeloma hücrelerinin (minimal rezidual hastalık-MRD) tedavisinde görüntüleme tedavinin etkinliği için çok büyük kolaylık gösterecektir. Wilms' tümör genini ekspresyon seviyelerinin real-time kuantitatif polimeraz zincir reaksiyonuyla ölçülmesi lösemi ve myelodisplastik sendromda minimal rezidual hastalık indikatörü olarak kullanımın yararı rapor edilmiştir. Bu çalışmanın hedefi WT1 ekspresyon seviyelerini ölçerek, bu genin ekspresyonu ve multiple myeloma arasındaki muhtemel ilişkinin teşhiste bulunmasıdır. Eğer bir ilişki bulunursa, WT1 geni diğer tanı faktörleriyle karşılaştırarak MRD markırı olarak kullanılabilmesidir. Bu çalışmada periferik kandaki WT1 ekspresyonunun seviyesini 50 yeni tanı almış multiple myeloma hastasında real-time light cycler kantitatif polimeraz zincir reaksiyonu kullanılarak ölçüldü. Normal WT1 gen kopya sayısı  $<23\mu\text{l}$  cDNA bulundu. Multiple myelomalı tüm hastalarda normal WT1-mRNA seviyesi bulundu. Bu çalışmaya göre WT1 ekspresyon analizlerinin multiple myelomalı hastaların teşhisinde rutin klinik uygulamalar için genetik markır olarak kullanımı uygun değildir.

## **ABSTRACT**

Monitoring patients with multiple myeloma during and after treatment for the presence of residual myeloma cells (minimal residual disease-MRD) has been shown to give a major insight into the effectiveness of treatment. It has been reported that Wilms tumor gene (WT1) expression levels measured by real time quantitative polymerase chain reaction was useful as an indicator of minimal residual disease in leukemia and myelodysplastic syndrome. The aim of this study was to measure levels of WT1 expression in order to find a possible association between the expression of this gene and multiple myeloma at diagnosis. If an association was found, the WT1 gene could be evaluated as an MRD marker by comparison with other prognostic factors. We investigated peripheral blood WT1 expression level measured by real time light cycler quantitative polymerase chain reaction in 50 newly diagnosed multiple myeloma patients. The normal WT1 gene copy number was found to be  $<23\mu\text{l}$  cDNA and all patients with myeloma were found to have normal WT1-mRNA levels. On this basis WT1 expression analysis is unlikely to be a useful genetic marker for routine clinical use in multiple myeloma patients at diagnosis.

## **GİRİŞ / AMAÇ VE KAPSAM**

Multiple Myelom' lu hastalarda tedavi öncesi ve tedavi sonrası Real-time PCR yöntemini kullanarak, WT1 geninin ekspresyonunun saptanması, saptanan bulguların karşılaştırılıp, hastalığın erken teşhisinde, takibinde veya tedavi başarısının izleminde kullanılabilirliğinin gerekliliğini tespit etmek. Aynı hastalarda 13 nolu kromozomun uzun kolunda bulunan RB1 genine konvansiyonel PCR yöntemi kullanarak 13q delesyona bakmak ve MM' lu hastalarda prognozla arasındaki ilişkinin açığa kavuşturulmasında literatüre değerli bilgilerin kazandırılmasıdır.

## GENEL BİLGİLER

Multiple myeloma (MM), terminal differansiye B hücrelerinin malignansisidir. Yıllık insidansı yaklaşık milyonda 40' dır yani ülkemizde yılda 2800 yeni vaka ortaya çıkmaktadır. İnsidansı yaşla artar (ortalama 65 yaş), 40 yaşın altında nadirdir, Afro karayiplilerde iki kat daha sıktır. Sadece radyasyon mazuriet kesin epidemiyolojik risk faktörü olarak tanımlanmıştır. Hastalık birkaç aydan 10 yıla kadar değişebilen süre ile oldukça heterojenöz sonuç göstermektedir. Konvansiyonel olarak kullanılan prognostik faktörler [serum b-2 mikroglobulin (b-2 m), albumin ve C reaktif protein (CRP) düzeyi, Durie- Salmon evreleme sistemi] hem kantitatif hem de her yerde kolaylıkla elde edilebileceğinden MM' li hastalarla ilgili klinik çalışmalarda oldukça kullanışlıdır. Bununla birlikte bu hastalardaki temel genetik anormallikleri gösteremezler.

Rpse et al. (1990) WT1 geninin 11p13 bölgesinde yerleştiğini saptamış ve Gessler et al. (1992) WT1 geninin genomik organizasyonunu tanımlamış, 10 exondan oluştuğunu bulmuştur. İki bölgedeki alternatif splicing in paterni, epidermal growth factor reseptör (EGFR), insulin-like growth factor-11 (IGF-II), IGF-I reseptör (IGFR-I) ve androgen receptor gibi büyüme faktörleri ve onların reseptörlerini transkripsiyon faktöründe, ayrıntılı olarak karakterize edilmiştir. wtWT1 proteininde, trans-aktivasyon domaini WT1' nin N-terminal kısmındadır ve onun transkripsiyonal aktivasyonu, bu transkripsiyon faktörlerinin hücre büyümesinin süprese etme yeteneği için kritiktir. Eğer bu trans-aktivasyonu damainini kodlayan dizide trWT1 transkripti yokken DNA bağlayan domaini mevcutsa, dominant negatif gibi etki gösterebilir ve direkt onkogeneze dahil olabilir. Wilm's tümöründe, WT1 geninde birçok farklı delesyonlar ve nokta mutasyonlar bildirilmiştir ki bununda mutasyona uğramış WT1 geninin ürünlerinin karsinogeneze rol oynayabileceğini düşündürmektedir. Yüksek ekspresyon düzeyleri akut lenfositik lösemiler, akut myeloid lösemiler ve kronik myelositik lösemilerde saptanmıştır. Ama ekspresyonu multiple myelomada araştırılmamıştır.

WT1 transkripsiyon faktörünün gerçekten lökomogeneze katkı mı yaptığı veya sadece immatur fetotipi gösterdiği immatür CD34+ progenitor hücrelerde normal olarak ekprese edildiği ve farklılaşmanın WT1 downregülasyonu ile ilişkili olduğunun bulunmasına kadar açık değildi. Tüm lösemili hastalarda lösemi hücrelerindeki WT1 ekspresyonunun yüksek olması kötü prognozla ilişkilidir. Üstelik WT1 ekspresyonunun yüksek olması kötü prognozla ilişkilidir. Üstelik WT1 ekspresyonu ve çoklu ilaç rezistansı bazı hematolojik malignensilerle ilişkilidir ki bu yüzden hematolojik malignensilerde kemorezistansta potansiyel diagnostik bir marker olabilir. WT1 ekspresyon düzeyinin lösemili vakaların sonucu ile direkt olarak korele olduğu düşünülmektedir. Bu yüzden transkripsiyon miktarının ve onun ana varyantlarının tam olarak değerlendirilmesi kritiktir ve hastalığın prognozu ve tanının doğruluğunu sağlar.

Anöploidi (flow cytometry ve fluorescence in situ hybridization (FISH) analizi ile değerlendirilebilir) esas olarak MM' lu tüm hastalarda olmaktadır. Akut myeloblastik I ve lenfoblastik lösemili hastalarda

sitogenetik çalışmaların prognostik önemi uzun süre önce tanımlanmıştır.MM da benzer çalışmaların sistematik değerlendirilmesi, temel olarak hastalığın düşük proliferatif doğası nedeniyle geri kalmıştır.Bu yüzden hastaların büyük çoğunluğu (yaklaşık 2/3 ü) non-myeloma hücrelerinin (normal hematopoietik) metafazlarını yansıtan non-informatif karyotipik bulgulara sahiptir.Konvansiyonel sitogenetik analizler diğer araştırmacılar tarafından HDT veya standart tedavilerle tedavi edilmiş MM' lı hastalarda kötü prognostik faktör olarak bildirilmiştir.Kötü prognozda del 13 arasında kesin bir ilişki henüz tespit edilmemiştir.Ama biz düşünmekteyizki del 13 olduğu zaman 13q14 bölgesindeki RB1 tümör süpresör geninin haplo yetersizliği ortaya çıkmaktadır.Gen ekspresyon profil teknolojisi,FISH ile del 13 saptanan hastalarda iki eksizyon tamir geninin önemli derecede down-regulasyon olduğunu tespit etmiştir.Böyle genlerin kaybı, genomik instabiliteye yol açabileceklerini ve böyle hastalarda kötü bir sonuç olacağını düşündürmektedir.

WT1 geninin akut myeloid lösemi (AML), akut lenfoblastik lösemi ve kronik myeloid lösemili hastalarda yüksek oranda eksprese olduğu bulunmuştur ve lösemik hücrelerin viabilitesinde anahtar bir rol oynadığı düşünülmektedir.WT1 geninin real time kantitatif zincir reaksiyonu ile kantitatif değerlendirilmesi son yıllarda değişik tip lösemilerde allojenik kemik iliği transplantasyonundan sonra minimal rezidüel hastalığın tespit edilmesinde marker olarak kullanılmıştır.Halbuki multiple astalarda WT1 geninin tedavi öncesi tedavi sonrası ekspresyonunun karşılaştırılıp hastalığın erken teşhisinde saptamak temel amacımızdır.Ayrıca 13. Kromozomun uzun kolunda bulunan RB1 genine konvansiyonel pcr yöntemiyle bakarak 13q delesyonu ile MM arasındaki ilişkinin açığa kavuşturulmasında literatüre değerli bilgilerin kazandırılması da ikinci amacımızdır.Ek yararlar olarak WT1 ekspresyonunun çoklu ilaç direnci ile ilişkisi hakkında şimdiye kadar bazı hematolojik malignensilerde bildirilmiş bilgilere ek veri sağlanabilecek ve ayrıca bu çalışmanın bir uzantısı olarak Insulin-like growth factor I-reseptörünün, myelomada otokrin büyüme sinyali döngüsünde etken olabileceği bulunmuştur ki (WT1 'in IGF1-reseptör geninin ekspresyonunun kontrolünde rol aldığı düşünülürse) bu bağlamda IGF1 reseptör geni ile ilgili çalışmalar planlanabilecektir.



## GEREÇ VE YÖNTEM

Eylül 2005 ve Mart 2007 tarihleri arasında yeni multiple myelom tanısı alan 50 yeni tanılı hasta (75 yaşından küçük, hiç terapi almamış) alındı. Multiple myeloma çalışmasına serum ve idrar protein elektroforezi ve immünoelektroforez, immünglobülinlerin serum seviyeleri kantitasyon, biyopside kemik iliği plazma hücre seviyesi, serum B2M, albümin, LDH ve CRP seviyeleri. Hastalar VAD ( vincristine 1 mg, günde 1; deksametazon 40mg/m<sup>2</sup>, günde 1-4; deksorubusin 50mg/m<sup>2</sup> günde 1) veya MP ( melphalan ve prednisolone) kemoterapi protokolüne göre tedavi edildi.

### GENETİK ANALİZ:

Periferik kan örnekleri (5ml, Etilendiamin tetra asetik asit) tanı anında her bir hastadan alındı. Toplamda 50 örnek analiz edildi. Altı (6) sağlıklı kişinin periferik kan örnekleri kontrol olarak kullanıldı.

Real –Time light cyclers kantitatif polimeraz zincir reaksiyonu (QRT-PCR)

RNA eldesi Qiagen RNA Blood mini kiti (Qiagen , Hilden, Germany) göre yapıldı. cDNA sentezi RT-PCR kitine göre yapıldı. Real-Time PCR Light Cycler Fast Start DNA SYBR Green kiti ile çalışıldı. Tüm primerler Search-LC ( Heidelberg, Germany) ‘ den alındı. Housekeeping gen olarak ABL geni kullanıldı. Her çalışma için pozitif ve negatif kontrol kullanıldı. Her bir örnekteki hedef molekül sayısı farklı hedeflerden elde edilen standart curve analizine göre yapıldı. Tüm örnekler ikişer kez çalışıldı.

## **BULGULAR**

WT1 ekspresyon seviyeleri negatif kontrol ve hasta grubu arasında anlamlı bir fark bulunamamıştır. (WT1 gen kopyaları < 23µl cDNA) MRD pozitif örneklerdeki WT1 seviyeleri bizim hastalarımıza göre oldukça yüksek bulundu. 50 örnekte de WT1 ekspresyonu tespit edilememiştir. İkincil hedefimiz WT1 geninin MRD takibinde marker olarak kullanılmasıdır.

## TARTIŖMA VE SONUÇ

WT1 geninin akut myeloid lösmi (AML), akute lenfoblastik lösemi ve kronik myeloid lösemili hastalarda yüksek oranda ekprese olduđu bulunmuştur ve lösemik hücrelerin viabilitesinde anahtar bir rol oynadıđı düşünölmektedir. WT1 geninin real time kantitatif zincir reaksiyonu ile kantitatif deđerlendirilmesi son yıllarda deđerşik tip lösemilerde allojenik kemik iliđi transplantasyonundan sonra minimal rezidüel hastalıđın tespit edilmesinde marker olarak kullanılmıştır. Fakat multiple myelomalı hastalarda WT1 gen ekspresyonu hakkında bir bilgi yoktur. Bu bađlamda MM'lı hastalarda WT1 geninin tedavi öncesi tedavi sonrası ekspresyonunun karşılaştıırılıp hastalıđın erken teşhisinde veya takibinde veya tedavi başarısının izleminde kullanılabilirliđinin tespitini real time PCR yöntemiyle saptamak temel amacımızdır.

## KAYNAKLAR

1-Ogawa H, Tamaki H, Ikegame K, et al. The usefulness of monitoring WT1 gene transcripts for the prediction and management of relapse following allogeneic stem cell transplantation in acute type leukemia. *Blood*. 2003;101:1698-1704.

2- Cilloni D, Gottardi E, Messa F, et al. Significant correlation between the degree of WT1 expression and the International Prognostic Scoring System Score in patients with myelodysplastic syndromes. *J Clin Oncol*. 2003;21:1988-1995.

3-Cilloni D, Saglio G. WT1 as a universal marker for minimal residual disease detection and quantification in myeloid leukemias and in myelodysplastic syndrome. *Acta Haematol*. 2004;112(1-2):79-84.

4-Uzunel M, Ringden O. Poor correlation of kinetics between BCR-ABL and WT1 transcript levels after allogeneic stem cell transplantation. *Bone Marrow Transplant*. 2004 Jan;33(1):47-52.

5-Galimberti S, Guerrini F, Carulli G, Faazi R, Palumbo GA, Morabito F, Petrini M. Significant co-expression of Wt1 and MDR1 genes in acute myeloid leukemia patients at diagnosis. *Eur J Haematol*. 2004 Jan;72(1):45-51.

6-Singer C, Samson D. Multiple myeloma. *Clin Med JRCPL* 2001;1:365-70.

7-Herrinton L J, Weiss NS, Olshan AF. Epidemiology of myeloma. In : Malpas JS, Bergsagel DE, RA, Anderson KC (eds). *Myeloma biology and management*. Oxford:Oxford University Press, 1998:150-86.

8-Athanasios B.-T. Fassas and Guido Tricot. Chromosome 13 Deletion/Hypodiploidy and Prognosis in Multiple Myeloma Patients. *Leukemia & Lymphoma*, June 2004 Vol. 45 (6), pp. 1083-1091.

9-Barlogie B., Epstein J., Selvanayagam, P. And Alexanian R. (1989) "Plasma cell myeloma –new biological insights and advances in therapy", *Blood*, 73, 865-879.

10-Kyle R.A. (1994) "Why better prognostic factors for multiple myeloma are needed", *Blood*, 83, 1713-1716.

11-Drach, J., Schuster, J., Nowotny, H., Angerler, J., Rosenthal, F., Fiegel, M., et al. (1995) ‘‘ Multiple myeloma: high incidence of chromosomal aneuploidy as detected by interphase fluorescence in situ hybridisation’’, *Cancer Research*, 55,3854-3859.

12-Latreille, J., Barlogie, B., Johnston, D., Drewinko, B. And Alexanian R. (1982) ‘‘ Ploidy and proliferative characteristics in monoclonal gammopathies ‘’, *Blood*, 59, 43-46.

13-WILMS TUMOR 1 GENE; WT1,OMIM, 607102.

14-Shaughnessy, J., Jacobson, J., Sawyer, J., McCoy, J., Fassas, A., Zhan, F., et al. (2003) ‘‘ Continuous absence of metaphase-defined cytogenetic abnormalities, especially of chromosome 13 and hypodiploidy, ensures long-term survival in multiple myeloma treated with Total Therapy I: interpretation in the context of global gene expression’’, *Blood*, 101, 3849-3856.

15-Fenaux, P., Preudhomme, C., Lai, J., Morel, P., Beuscart, R. And Bauters, F. (1989) ‘‘Cytogenetics and their prognostic value in de novo acute myeloid leukaemia: a report on 283 cases ‘’, *British journal of Haematology*, 73,61-67.

16-Secker-Walker, L.M. (1990) ‘‘Prognostic and biological importance of chromosome findings in acute lymphoblastic leukemia ‘’, *Cancer Genetics Cytogenetics*,49, 1-13.

17-Shaughnessy J. Jr., Tian, E., Sawyer, J., McCoy, J., Tricot, G., Jacobson, J., et al.(2003) ‘‘ Prognostic impact of cytogenetic and interphase fluorescence in situ hybridization-defined chromosome 13 deletion in multiple myeloma: early result of total therapy II’’, *British Journal of Haematology*, 100, 44-52.

18-Seong, C., Delasalle, K., Hayes, K., Weber, D., Dimopoulos, M., Swankowski, J., et al. (1998) Prognostic value of cytogenetics in multiple myeloma. *British Journal of Haematology*, 101, 189-194.

19-Weh, H.J., Gutensohn, K., Selbach, J., Kruse, R., Wacker-Backhaus, G., Seeger, D., et al. (1993) ‘‘ Karyotype in multiple myeloma and plasma cell leukemia’’, *European Journal of Cancer*, 29A, 1269-1273.

20-La, J.L., Zandecki, M., Mary, J.Y., Bernardi, P., Izydorzyc, V., Flactif, M., et al. (1995) ‘‘Improved cytogenetics in multiple myeloma and: a study of 151 patients including 117 patients at diagnosis’’, *Blood*, 85, 2490-2497.

## **EKLER**

**1-**Makale ekte sunulmuştur.

# **Expression of WT1 gene in multiple myeloma patients at diagnosis: is WT1 gene expression a useful marker in multiple myeloma?**

Cetin Saatci<sup>1</sup>, A. Okay Caglayan<sup>1</sup>, Ismail Kocyigit<sup>2</sup>, Hilal Akalin<sup>1</sup>, Leyla G. Kaynar<sup>2</sup>,Fevzi Altuntas<sup>2</sup>, Bulent Eser<sup>2</sup>, Muzaffer Demir<sup>3</sup>, Mustafa Cetin<sup>2</sup> and Yusuf Ozkul<sup>1</sup>

<sup>1</sup>Department of Medical Genetics,

<sup>2</sup>Department of Hematology, Medical Faculty, Erciyes University, Kayseri and

<sup>3</sup>Department of Hematology, Medical Faculty, Trakya University, Edirne, Turkey

Monitoring patients with multiple myeloma during and after treatment for the presence of residual myeloma cells (minimal residual disease – MRD) has been shown to give a major insight into the effectiveness of treatment. It has been reported that Wilms' tumor gene (WT1) expression levels measured by real-time quantitative polymerase chain reaction was useful as an indicator of minimal residual disease in leukemia and myelodysplastic syndrome. The aim of this study was to measure levels of WT1 expression, in order to find a possible association between the expression of this gene and multiple myeloma at diagnosis. If an association was found, the WT1 gene could be evaluated as an MRD marker by comparison with other prognostic factors. We investigated peripheral blood WT1 expression level measured by real-time light cycler quantitative polymerase chain reaction in 50 newly diagnosed multiple myeloma patients. The normal WT1 gene copy number was found to be ,23/ml cDNA and all patients with myeloma were found to have normal WT1-mRNA levels. On this basis WT1 expression analyses is unlikely to be a useful genetic marker for routine clinical use in multiple myeloma patients at diagnosis.

Keywords: Expression, multiple myeloma, WT1

## **Introduction**

Multiple myeloma (MM) is a tumor of bone marrow plasma cells. It accounts for 10% of hematologic malignancies and 1% of all cancer deaths.<sup>1</sup> Incidence increases with age (median 65 years), it is rare under 40, and twice as common in Afro-Caribbeans. The only clearly defined epidemiological risk factor is radiation exposure.<sup>2</sup> Several adverse prognostic factors including paraprotein levels, Hb count, serum calcium level, lytic bone lesions, and renal function have been identified. These factors are combined in the widely used Durie-Salmon staging system.<sup>3</sup> Renal impairment is a risk factor only because of its association with high tumour burden.<sup>4</sup> Additionally, the international prognostic index (IPI) consisting of beta-2 microglobulin and serum albumin level is also used as a prognostic factor. However, the parameters used do not address the fundamental genetic abnormalities of the disease. Deletion of 13q is an important adverse prognostic factor<sup>5</sup> and the presence of hypodiploidy is strongly associated with poor prognosis in MM.<sup>6</sup> In contrast to the acute myeloblastic and lymphoblastic leukemias<sup>7,8</sup> where the prognostic importance of cytogenetic studies has long been recognized, the systematic evaluation of similar studies in MM has lagged behind, mainly because of the low proliferative nature of the disease. Therefore, the majority of patients (approximately two-thirds at presentation) have non-informative karyotypic studies which reflects the analysis of only the metaphases of (normal hematopoietic) non-myeloma cells.

Correspondence to: A. Okay Caglayan, MD Erciyes University, Medical Faculty, Department of Medical Genetics, Talas Caddesi, 38039, Kayseri, Turkey

E-mail: aocaglayan@erciyes.edu.tr

© W. S. Maney & Son Ltd 2010

Received 5 April 2009; accepted 17 July 2009

DOI 10.1179/102453310X12583347009496 Hematology 2010 VOL 15 NO 1 39



In recent years, the use of Wilms' tumor gene as a 'panleukemic' marker has been investigated in various studies.<sup>9–11</sup> WT1 encodes a transcription factor involved in the pathogenesis of Wilms' tumor. Although the role of WT1 in tumorigenesis has not been well defined, further analyses of the gene have shown that the level of expression is high in most patients with acute leukemia and CML.<sup>12,13</sup> Longterm monitoring of WT1 levels has been used to detect early relapse and predict the prognosis after chemotherapy or allogeneic stem cell transplantation.<sup>14–16</sup> An association between WT1 expression and relapse has been found a number of studies.<sup>17–19</sup> However, there is very little data concerning the monitoring of WT1 levels in myeloma patients and its effect on prognosis. Our primary goal was to evaluate peripheral blood (PB) WT1 level as genetic marker at diagnosis and the secondary goal was to determine the association with the WT1 gene as an MRD marker by comparing the other prognostic factors.

## **Methods**

**Patients** Between September 2005 and March 2007, 50 newly diagnosed patients with symptomatic MM (75 years or younger, no prior therapy) were enrolled in this prospective study (Table 1). Written informed consent had been obtained from all participants in keeping with institutional and National Cancer Institute guidelines. Multiple myeloma work-up included analysis of serum and urine protein electrophoresis and immunoelectrophoresis, quantitation of serum levels of immunoglobulins, bone marrow plasma cell level on biopsy and aspirate samples, serum B2M, albumin, LDH and CRP levels. Baseline MRI examinations were performed of the axial skeleton (head, shoulders, sternum, spine and pelvis). 20 Patients were treated with VAD (vincristine 1 mg, day 1; dexamethasone 40 mg/m<sup>2</sup>, days 1–4; doxorubicin 50 mg/m<sup>2</sup>, day 1) or MP (melphalan and prednisolone) chemotherapy protocol.<sup>21</sup>

### **Genetic analysis**

Peripheral blood samples (5 ml, ethylene diamine tetra acetate) were drawn from each patient at diagnosis. A total of 50 samples were analyzed. PB samples from six healthy volunteers were used as controls. Real-time light cycler quantitative polymerase chain reaction (QRT-PCR) RNA preparation and reverse transcription: The total RNA was isolated from 1 ml of buffy coat, obtained through centrifugation (400 g for 10 min). The buffy coat was transferred to 6 ml of red cell lysis buffer included in the Qiagen RNA Blood Mini-kit (Qiagen, Hilden,

Germany). RNA extraction was then performed according to the manufacturer's instructions. The cDNA was prepared using first strand cDNA synthesis kit for RT-PCR according to the manufacturer's instructions. Real-time PCR was performed with the Light Cycler Fast Start DNA SYBR Green kit. All primers were obtained from Search-LC (Heidelberg, Germany). The calculated number of specific transcripts was normalized to the housekeeping gene ABL and presented as adjusted transcripts/ml cDNA. We used negative and positive controls of each run. The number of target molecules in each unknown sample was calculated automatically using the standard curves from each different target. All samples were run in duplicate and a negative control was included in each run.

## Results

The WT1 expression levels were not significantly different between negative control and the patient group (WT1 gene copies ,23/ml cDNA). In addition, WT1 levels in MRD positive samples were significantly higher than in our patients (P,0.05). Because WT1 expression was not detected in 50 samples, the secondary goal to determine the association with the WT1 gene as an MRD marker by comparing the other prognostic factors was not performed.

Table 1 Patients clinical and laboratory characteristics

Number of patients	50
Median age (range), years	57.5 (30–81)
Males females, n	32 male, 18 female
M component isotype, n	50
IgG	31
IgA	12
Non-secretuar	7
Light chain, n	50
Kappa	20
Lambda	14
Non k, non l	9
Stage (Durie-Salmon)	50
I	9
II	22
III	19

Median B2MG level (range), mg/l	11.85 (2.6–24.3)
Median CRP level (range), mg/l	18.95 (2.3–152)
Median LDH level (range), U/l	145 (100–190)
Median Hb level (range), mg/dl	9.8 (6 . 8–13.7)
Median BMPC (range),	% 55% (20–95%)

B2MG: b2-microglobulin; CRP: C-reactive protein; BMPC: bone marrow plasma cells; LDH: lactat dehydrogenase.

## Discussion

The clinical course of patients with multiple myeloma is highly variable. A large part of newly diagnosed cases of MM have a normal karyotype and abnormalities are more often seen in advanced disease that is associated with an increased proliferative activity of malignant cells. FISH testing for specific chromosomal abnormalities in patients with normal chromosome studies has identified an abnormal clone in 45% of cases.<sup>22</sup> Deletions of 13q14, 17p13, and 11q23 are all associated with poor prognostic factor.<sup>23–25</sup> 14q32 translocations involving chromosomes 4 and 16 have also been identified.<sup>8</sup> Additional common cytogenetic changes can be seen as hyper- or hypodiploidy and abnormalities involving the chromosome 1q. Also further genetic instability is initiated through alteration of proliferation or apoptosis pathways. Pathways with identified mutations include the nuclear factor-kB pathway, ras and p53. Sensitive MRD assays are essential for detection of a threatening relapse and the early start of immunotherapeutic interventions. However, current MRD assays are heterogeneous and disease specific approaches are sometimes needed. Therefore, the idea of using WT1 as a MRD marker was an obvious area of study in myeloma patients. Hatta et al. examined the association of clinical parameters and WT1 expression in bone marrow for 17 newly diagnosed multiple myeloma patients and found WT1 transcripts increased when clinical factors deteriorate, including the stage, amount of M protein, Hb, platelet count, blood urea nitrogen, creatinine, serum alkaline phosphatase, calcium, beta2-microglobulin, thymidine kinase activity, and C-reactive protein.<sup>26</sup> In contrast, an association between WT1 expression and multiple myeloma was not shown in present study. This difference may be partly explained by differences in sensitivities, the use of qualitative versus quantitative analyses and sample type. The concrete role of WT1 in hematopoiesis and leukemogenesis still remains unclear. It is reported that WT1 expression levels in PB samples by quantitative RTPCR were

significantly lower when compared to bone marrow samples in healthy individuals and the WT1 expression level was quite low and sometimes undetectable by quantitative RT-PCR.<sup>27</sup> On the other hand, Inoue et al. reported that WT1 expression in BM was one log of magnitude higher than in PB.<sup>14</sup> In present study, a comparison of the range of WT1 levels in patients and MRD positive samples showed that all of the patients are within the same range as the MRD negative ones. Therefore, WT1 analysis in BM may be more sensitive than PB in myeloma patients. In conclusion, there is a constitutive low expression of WT1 in myeloma patients in peripheral blood and WT1 expression analysis may not be a practical genetic marker for routine clinical use in myeloma patients at diagnosis.

## References

1. Kyle RA, Rajkumar SV. Multiple myeloma. *N Engl J Med* 2004; 351: 1860–1873.
2. Herrinton LJ, Weiss NS, Olshan AF. Epidemiology of myeloma. In: Malpas JS, Bergsagel DE, Kyle RA, Anderson KC. (eds) *Myeloma, biology and management*. Oxford: Oxford University Press, 1998; 150–186.
3. Durie BG, Salmon SE. A clinical staging system for multiple myeloma. Correlation of measured myeloma cell mass with presenting clinical features, response to treatment, and survival. *Cancer* 1975; 36: 842–854.
4. Iggo N. Management of renal complications. In: Malpas JS, Bergsagel DE, Kyle RA, Anderson KC. (eds) *Myeloma, biology and management*. Oxford: Oxford University Press, 1998; 381–401.
5. Tricot G, Barlogie B, Jagannath S et al. Poor prognosis in multiple myeloma is associated only with partial or complete deletion of chromosome 13 or abnormalities involving 11q and not with other karyotype abnormalities. *Blood* 1995; 86: 4250–4256.
6. Smadja NV, Bastard C, Brigaudeau C, Leroux D, Fruchart C. Hypodiploidy is a major prognostic factor in multiple myeloma. *Blood* 2001; 98: 2229–2238.

7. Fenaux P, Preudhomme C, Lai JL et al. Cytogenetics and their prognostic value in de novo acute myeloid leukaemia: a report on 283 cases. *Br J Haematol* 1989; 73: 61–67.
8. Secker-Walker LM. Prognostic and biological importance of chromosome findings in acute lymphoblastic leukemia. *Cancer Genet Cytogenet* 1990; 49: 1–13.
- 9 Inoue K, Sugiyama H, Ogawa H et al. WT1 as a new prognostic factor and a new marker for the detection of minimal residual disease in acute leukemia. *Blood* 1994; 84: 3071–3079.
10. Bergmann L, Miething C, Maurer U et al. High levels of Wilms' tumor gene (WT1) mRNA in acute myeloid leukemias are associated with a worse long-term outcome. *Blood* 1997; 90: 1217–1225.
11. Kreuzer KA, Saborowski A, Lupberger J et al. Fluorescent 50-exonuclease assay for the absolute quantification of Wilms' tumor gene (WT1) mRNA: implications for monitoring human leukaemias. *Br J Haematol* 2001; 114: 313–318.
12. Miwa H, Beran M, Saunders GF. Expression of the Wilms' tumor gene (WT1) in human leukemias. *Leukemia* 1992; 6: 405–409.
13. Miyagi T, Ahuja H, Kubota T, Kubonishi I, Koeffler HP, Miyoshi I. Expression of the candidate Wilms' tumor gene, WT1, in human leukemia cells. *Leukemia* 1993; 7: 970–977.
14. Inoue K, Ogawa H, Yamagami T et al. Long-term follow-up of minimal residual disease in leukemia patients by monitoring WT1 (Wilms tumor gene) expression levels. *Blood* 1996; 88: 2267–2278.
- 15 Kletzel M, Olzewski M, Huang W, Chou PM. Utility of WT1 as a reliable tool for the detection of minimal residual disease in children with leukemia. *Pediatr Dev Pathol* 2002; 5: 269–275.

16. Ogawa H, Tamaki H, Ikegame K et al. The usefulness of monitoring WT1 gene transcripts for the prediction and management of relapse following allogeneic stem cell transplantation in acute type leukemia. *Blood* 2003; 101: 1698–1704.
17. Gaiger A, Schmid D, Heinze G et al. Detection of the WT1 transcript by RT-PCR in complete remission has no prognostic relevance in de novo acute myeloid leukemia. *Leukemia* 1998; 12: 1886–1894.
18. Elmaagacli AH, Beelen DW, Trenchel R, Schaefer UW. The detection of wt-1 transcripts is not associated with an increased leukemic relapse rate in patients with acute leukemia after allogeneic bone marrow or peripheral blood stem cell transplantation. *Bone Marrow Transplant* 2000; 25: 91–96.
19. Schmid D, Heinze G, Linnerth B et al. Prognostic significance of WT1 gene expression at diagnosis in adult de novo acute myeloid leukemia. *Leukemia* 1997; 11: 639–643.  
Saatci et al. WT1 gene expression in multiple myeloma *Hematology* 2010 VOL 15 NO 1 41
20. Walker R, Barlogie B, Haessler J et al. Magnetic resonance imaging in multiple myeloma: diagnostic and clinical implications. *JCO* 2007; 25: 1121–1128.
21. Tribalto M, Amadori S, Cudillo L et al. Autologous peripheral blood stem cell transplantation as first line treatment of multiple myeloma: an Italian Multicenter Study. *Haematologica* 2000; 85: 52–58.
22. Konigsberg R, Zojer N, Ackerman J et al. Predictive role of interphase cytogenetics for survival of patients with multiple myeloma. *J Clin Oncol* 2000; 4: 804–812.
23. Harrison CJ, Mazzullo H, Cheung KL et al. Cytogenetics of multiple myeloma: interpretation of fluorescence in situ hybridization results. *Br J Haematol* 2003; 120: 944–952.
24. Seong C, Delasalle K, Hayes K et al. Prognostic value of cytogenetics in multiple myeloma. *Br J Haematol* 1998; 101: 189–194.

25. Fonseca R, Blood E, Rue M et al. Clinical and biological implications of recurrent genomic aberrations in myeloma. *Blood* 2003; 101: 4569–4575.
26. Hatta Y, Takeuchi T, Saitoh T et al. WT1 expression level and clinical factors in multiple myeloma. *J Exp Clin Cancer Res* 2005; 24: 595–599.
27. Lin F, Goldman JM, Cross NC. A comparison of the sensitivity of blood and bone marrow for the detection of minimal residual disease in chronic myeloid leukaemia. *Br J Haematol* 1994; 86: 683–685. Saatci et al. WT1 gene expression in multiple myeloma 42 *Hematology* 2010 VOL 15 NO 1